

Efficient Preparation of Aminoxyacyl Amides, Aminoxy Hybrid Peptides, and α-Aminoxy Peptides

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Received July 24, 2009



N-(Pg- α -aminoxy acids) **1a**-**g** are converted to *N*-(Pg- α -aminoxyacyl)benzotriazoles **2a**-**g**, which react under mild conditions with amines, α -amino acids/ α -dipeptides, and α -aminoxy acids to give aminoxyacyl amides **3a**-**g**, (**3e** + **3e**'), and (**3g** + **3g**'), aminoxy hybrid peptides **4a**-**h**, (**4a** + **4a**'), **6a**-**d**, **9a**-**e**, (**9a** + **9a**'), and (**9b** + **9b**'), and α -aminoxy peptides **10a**,**b** in good yields without racemization.

Introduction

In peptidomimetic foldamer chemistry, β -peptides have been applied widely in biomolecular design.¹ Unlike α -peptides, short β -peptides can fold into well-defined secondary structures, such as helices, sheets, and turns. Since β -peptides have excellent stability toward proteases,² they are widely used as backbone-modified amino acids in drug design. α -Aminoxy acids are analogues of β -amino acids in which the β -carbon atom is replaced by an oxygen atom. An α -aminoxy acid is more rigid than its corresponding β -amino acid,^{3a} and aminoxy

8690 J. Org. Chem. 2009, 74, 8690–8694

amide bonds are resistant to enzymatic degradation; therefore, α -aminoxy acids have been explored as peptidomimetics.^{3b}

 α -Aminoxy peptides have attracted considerable interest as novel foldamers⁴ because of their unusual conformations and interesting bioactivities.⁵ Aminoxy peptides may feature strong intramolecular hydrogen bonds between adjacent residues in peptidomimetic foldamers.⁶ For example, peptides consisting of α -aminoxy acids can possess eight-membered-ring intramolecular hydrogen bonds (α N–O turns),⁷ and peptides consisting of β -aminoxy acids can possess ninemembered-ring intramolecular hydrogen bonds (β N–O turns).^{5e} Oligomers of homochiral α - or β -aminoxy acids can form helical structures consisting of consecutive N-O turns (1.88 and 1.79 helices, respectively).⁸ β -Sugar aminoxy peptides exhibited rigid ribbon-like secondary structures composed of 5/7 bifurcated intramolecular hydrogen bonds.^{6a} Hybrid peptides of an α -aminoxy acid provided robust 12/10-mixed helices.⁹

Published on Web 10/15/2009

DOI: 10.1021/jo901612j © 2009 American Chemical Society

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Peptides containing a aminoxy acids are good receptors for anions because of the acidity of their aminoxy amide protons.¹⁰ A compound derived from an α -aminoxy acid has been used as an effective chemical shift reagent for measuring the ee of carboxylic acids;^{11a} another compound derived from an α -aminoxy acid forms chloride channels to mediate chloride ion transportation across cell membranes.11b

Published methods for the preparation of aminoxy acid derivatives and aminoxy peptides include (i) combinations of coupling reagents such as Bop-HOBt-NEM, HBTU-HOBt-NEM, DIC-HOAt;¹² EDCl-HOBt, EDCl-HOAt;^{6a} TBTU/ HOBt/DIEA;¹³ HOBt, BOP, DIEA;¹⁴ *i*BuOCOCl/NMM;^{5d} DIC/HOBt;¹⁵ (ii) activated esters;^{16,17} and (iii) α -amino diazoketone.¹⁸ These methods often involve longer reaction times¹³ and N-diacylation products¹⁶ and give low yields.¹⁵ Hence, there is a need for a mild and efficient general method to prepare aminoxyacyl amides, aminoxy hybrid peptides, and aminoxy peptides.

N-Acylbenzotriazoles are stable, mostly crystalline compounds and easy to handle. These N-acylbenzotriazoles are advantageous for N-, O-, C-, and S-acylation, 19 especially where the corresponding acid chlorides are unstable or difficult to prepare.^{191,m} N-Fmoc-(α -aminoacyl)benzotriazoles and their Boc- and Cbz- analogues enabled the preparation of chiral di-, tri-, and tetrapeptides in good yields from natural amino acids in solution phase with complete retention of chirality.19b,20 Recently, we have also prepared peptide alcohols in good yields using N-protected (α -aminoacyl)benzotriazoles and N-protected $(\alpha$ -dipeptidoyl)benzotriazoles.²¹ Herein, we describe a new method for the preparation of aminoxyacyl amides, aminoxy hybrid peptides, and aminoxy peptides by using N-protected $(\alpha$ -aminoxyacyl)benzotriazoles 2.

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Results and Discussion

Preparation of N-Pg(α -aminoxyacyl)benzotriazole 2a-g. *N*-Protected (α -aminoxy) acids **1b**-**g** were synthesized from either corresponding α -bromocarboxylic acids or α -hydroxycarboxylic acids and were well characterized by ¹H NMR, ¹³C NMR, and elemental analysis. The complete reaction procedure and the characterization data of N-protected (α aminoxy)acids 1b-g are given in the Supporting Information. N-Pg(α -aminoxy) acid **1a** was obtained from a commercial source. N-Pg(α -aminoxyacyl)benzotriazoles 2a-g have been prepared by treatment of N-Pg(α -aminoxy) acids 1a-g with 4 equiv of 1*H*-benzotriazole and 1 equiv of thionyl chloride in THF at room temperature in 56-89% yields (Scheme 1, Table 1). Products 2a-g were characterized by ¹H NMR, ¹³C NMR, and elemental analysis.

SCHEME 1. Preparation of N-Pg(α -aminoxyacyl)benzotriazoles 2a-g



TABLE 1. Preparation of N-Pg(α -aminoxyacvl)benzotriazoles 2a-g

				-
entry	Pg: protecting group	R	2 , yield ^{<i>a</i>} (%)	mp (°C)
1	<i>tert</i> -butoxycarbonyl (Boc)	Н	2a , 67	114-115
2	phthalimide (Phth)	Н	2b , 75	155-157
3	phthalimide (Phth)	Me	2c , 56	145-147
4	benzyloxycarbonyl (Cbz)	Н	2d, 66	86-87
5	Cbz	Me	2e , 58	oil
6	Cbz	$CH(CH_3)_2$	2f , 89	oil
7	Cbz	Ph	2g , 77	oil
^a Isolated yield.				

Preparation of N-Pg(α -aminoxyacyl)amides 3a-g, (3e+3e'), and (3g+3g'). α -Aminoxyacylamides also exhibit intramolecular hydrogen bonds between adjacent residues (α N-O turns) in peptidomimetic foldamers.^{6b,7a} N-Pg(α aminoxyacyl) amides 3a-g, (3e+3e'), and (3g+3g') were obtained by reaction between N-Pg(α -aminoxyacyl)benzotriazoles 2a,b,d-f and the corresponding amines in THF at room temperature in the presence of triethylamine in 38-78% yields (Scheme 2, Table 2). Products 3a-g, (3e+3e'), and (3g+3g') were characterized by ¹H NMR, ¹³C NMR, and elemental analysis. Retention of enantiopurity of product 3g was confirmed by chiral HPLC using a Whelk-O1 column (with detection at 254 nm, a flow rate of 1.0 mL/min, and MeOH as the eluting solvent). The diastereomer 3g showed a single retention-time peak in chiral HPLC at 3.46, while its corresponding diastereomeric mixture (3g+3g') showed two peaks at 3.46 and 5.68.

SCHEME 2. Preparation of N-Pg(α -aminoxyacyl)amides 3a-g, (3e+3e'), and (3g+3g')



Synthesis of *a*-Aminoxy Hybrid Peptides. *a*-Aminoxy hybrid peptides are defined as including at least one

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 TABLE 2.
 Preparation of N-(Pg)-aminoxy Acid Amides 3a-g, (3e+3e'), and (3g+3g')

entry	2	amine	3 , yield ^{<i>a</i>} (%)	mp (°C)
1	Boc-AOGly-Bt, 2a	isopropylamine	3a , 74	67-68
2	Boc-AOGly-Bt, 2a	<i>c</i> -hexylamine	3b , 62	137-139
3	Cbz-AOGly-Bt, 2d	<i>c</i> -hexylamine	3c ,78	69-70
4	Phth-AOGly-Bt, 2b	<i>p</i> -methoxyaniline	3d , 38	210-211
5	Cbz-AOGly-Bt, 2d	L-2-methylbenzylamine	3e , 67	oil
6	Cbz-AOGly-Bt, 2d	DL-2-methylbenzylamine	(3e+3e'), 73	oil
7	Cbz-L-AOÁla-Bt, 2e	<i>p</i> -methoxyaniline	3f , 56	31-32
8	Cbz-L-AOVal-Bt, 2f	L-2-methylbenzylamine	3 g, 67	96-97
9	Cbz-L-AOVal-Bt, 2f	DL-2-methylbenzylamine	(3g + 3g '), 58	oil
^a Isolated vie	eld ${}^{b}AO$ stands for aminoxy in the nom	enclature for aminoxy compounds through	out the paper	

TABLE 3. α -AO- α -hybrid Dipentides 4a-h and (4a+4a')

entry	2	amino acid	product 4 , yield ^{a} (%)	mp (°C)
1	Cbz-AOGly-Bt, 2d	L-Phe-OH	Cbz-AOGly-L-Phe-OH, 4a, 51	oil
2	Cbz-AOGly-Bt, 2d	DL-Phe-OH	Cbz-AOGly-DL-Phe-OH, $(4a+4a')$, 56	oil
3	Cbz-L-AOAla-Bt, 2e	L-Phe-OH	Cbz-L-AOAla-L-Phe-OH, 4b, 61	33-34
4	Cbz-L-AOAla-Bt, 2e	L-Trp-OH	Cbz-L-AOAla-L-Trp-OH, 4c, 72	128-130
5	Cbz-L-AOAla-Bt, 2e	L-Leu-OH	Cbz-L-AOAla-L-Leu-OH, 4d, 78	oil
6	Cbz-L-AOVal-Bt, 2f	L-Phe-OH	Cbz-L-AOVal-L-Phe-OH, 4e, 69	126-127
7	Cbz-L-AOVal-Bt, 2f	L-Trp-OH	Cbz-L-AOVal-L-Trp-OH, 4f, 66	26-27
8	Cbz-AOGly-Bt, 2d	L-Cys-OH	Cbz-AOGly-L-Cys-OH, 4g, 66	oil
9	Cbz-L-AOAla-Bt, 2e	L-Cys-OH	Cbz-L-AOVal-L-Cys-OH, 4h, 61	oil
^a Isolated yi	eld.			

TABLE 4. Preparation of α-AO-α,α-hybrid Tripeptides 6a-d

entry	2	dipeptide, 5	product 6 , yield ^{a} (%)	mp (°C)
1	Cbz-AOGly-Bt, 2d	Gly-L-Phe-OH	Cbz-AOGly-Gly-L-Phe-OH, 6a, 81	oil
2	Cbz-AOGly-Bt, 2d	Gly-L-Leu-OH	Cbz-AOGly-Gly-L-Leu-OH, 6b, 70	oil
3	Cbz-L-AOAla-Bt, 2e	Gly-L-Phe-OH	Cbz-L-AOAla-Gly-L-Phe-OH, 6c, 50	63-64
4	Cbz-L-AOVal-Bt, 2f	Gly-L-Phe-OH	Cbz-L-AOVal-Gly-L-Phe-OH, 6d, 67	123-124
^a Isolated	vield.			

 α -aminoxy acid residue and at least one natural amino acid residue. We have prepared α -AO- α -hybrid dipeptides **4a**-**h** and (**4a**+**4a**'), α -AO- α , α -hybrid tripeptides **6a**-**d**, and α , α -AO-hybrid dipeptides **9a**-**d** and (**9a**+**9a**').

Preparation of α-AO-α-hybrid dipeptides 4a-h and (4a+4a'). α-AO-α-Hybrid dipeptides 4a-h and (4a+4a') were prepared by treatment of *N*-(Pg-aminoxyacyl)benzo-triazoles 2d-f and the corresponding amino acids in CH₃-CN-H₂O in the presence of triethylamine at room temperature in 51-78% yields (Scheme 3, Table 3). Products 4a-h were characterized by ¹H NMR, ¹³C NMR, and elemental analysis. Retention of enantiopurity of α-AO-α-hybrid dipeptides 4a and (4a+4a') was supported by chiral HPLC analysis using a Chirobiotic T column (detection at 254 nm, flow rate 1.0 mL/min, and MeOH as eluent). α-AO-α-hybrid dipeptide 4a showed a single retention-time peak in chiral HPLC analysis at 3.15, while the corresponding enantiomeric mixture (4a+4a') showed two peaks at 2.89 and 3.69.

SCHEME 3. a-AO-a-hybrid Dipeptides 4a-h and (4a+4a')



Preparation of α-AO-α,α-hybrid tripeptides 6a-d. α-AOα,α-hybrid tripeptides 6a-d were prepared by treatment of *N*-Pg(α-aminoxyacyl)benzotriazoles 2d-f with unprotected dipeptides **5a,b** in CH₃CN-H₂O in the presence of triethylamine in 50-81% yields (Scheme 4, Table 4).

SCHEME 4. Preparation of α -AO- α , α -hybrid Tripeptides 6a-d



Preparation of α, α -AO-hybrid Dipeptides 9a-e, (9a+9a'), and (9b+9b'). α, α -AO-hybrid dipeptides 9a-e, (9a+9a'), and (9b+9b') were obtained by treatment of *N*-Pg(α -aminoacyl)benzotriazole 7a-c and the corresponding aminoxy acids 8a,b (details about preparation of compounds 8a,b are given in the Supporting Information) in CH₃CN-H₂O (3:1) in the presence of triethylamine at room temperature in 44-90% yields (Scheme 5, Table 5). All products 9a-e, (9a+9a'), and (9b+9b') were characterized by ¹H NMR, ¹³C NMR, and elemental analysis. The enantiomeric purity of

SCHEME 5. Preparation of α,α -AO-hybrid Dipeptides 9a-e, (9a+9a'), and (9b+9b')



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TABLE 5. Preparation of α , α -AO-hybrid Dipeptides 9a-e, (9a+9a'), and (9b+b')

entry	7	8	product 9	yield ^{a} (%)	mp (°C)
1	Cbz-L-Ala-Bt, 7a	AOGly-OH, 8a	Cbz-L-Ala-AOGly-OH, 9a	61	24-25
2	Cbz-DL-Ala-Bt, $(7a+7a')$	AOGly-OH, 8a	Cbz-DL-Ala-AOGly-OH, (9a+9a')	71	24-25
3	Cbz-L-Phe-Bt, 7b	AOGly-OH, 8a	Cbz-L-Phe-AOGly-OH, 9b	90	121-122
4	Cbz-DL-Phe-Bt, $(7b+7b')$	AOGly-OH, 8a	Cbz-DL-Phe-AOGly-OH, (9b+9b')	72	145-146
5	Cbz-L-Met-Bt, 7c	AOGly-OH, 8a	Cbz-L-Met-AOGly-OH, 9c	81	87-88
6	Cbz-L-Phe-Bt, 7b	L-AOAla-OH, 8b	Cbz-L-Phe-L-AOAla-OH, 9d	44	38-39
7	Cbz-L-Met-Bt, 7c	L-AOAla-OH, 8b	Cbz-L-Met-L-AOAla-OH, 9e	73	127-128
^a Isolate	ed yield.				

TABLE 6. Preparation of α-Aminoxy Dipeptides 10a and 10b

entry	2	8	product 10 , yield ^{a} (%)
1	Cbz-L-AOVal-Bt, 2f	AOGly-OH, 8a	Cbz-L-AOVal-AOGly-OH, 10a, 58
2	Cbz-L-AOMan-Bt, 2g	L-AOAla-OH, 8b	Cbz-L-AOMan-L-AOAla-OH, 10b, 52
^a Isolated vield.			

 α,α -AO-hybrid dipeptides **9b** and (**9b**+**9b**') was supported by chiral HPLC analysis using a Chirobiotic T column (detection at 254 nm, flow rate 0.6 mL/min, and MeOH-H₂O (7:3) as eluent). α,α -AO-hybrid dipeptide **9b** showed a single peak in HPLC analysis at 4.74, while the corresponding enantiomeric mixture (**9b**+**9b**') showed two peaks at 4.10 and 5.32.

Preparation of α-Aminoxy Dipeptides 10a,b. α-Aminoxy dipeptides **10a,b** were prepared in 55–58% yields by reaction of *N*-(Pg-aminoxyacyl)benzotriazoles **2f**,g with the corresponding aminoxy acids **8a,b** in CH₃CN-H₂O (3:1) in the presence of triethylamine at ambient temperature (Scheme 6, Table 6). Products **10a,b** were characterized by ¹H NMR, ¹³C NMR, and elemental analysis.

SCHEME 6. Preparation of α-Aminoxy Dipeptides 10a and 10b



In comparison with literature methods, our method has the following advantages: (i) utilization of milder reaction conditions, (ii) shorter reaction times, (iii) better yields, (iv) use of inexpensive, easily synthesizable N-(Pg- α -aminoxyacyl)benzotriazole reagents for peptide coupling, (v) coupling without protecting the carboxylic acid group, (vi) no N-diacylation products, and (vii) avoids racemization.

Conclusions

In conclusion, a mild and an efficient general method for the preparation of α -aminoxyacylamides, α -aminoxy hybrid peptides, and α -aminoxy peptides has been developed by reacting *N*-(Pg- α -aminoxyacyl)benzotriazoles with amines, α -amino acids, peptides, and α -aminoxy acids. All of the α -aminoxy derivatives were obtained under mild reaction conditions in good yields and with no racemization.

Experimental Section

Melting points are uncorrected. ¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded on a 300 MHz apparatus in CDCl₃ or DMSO- d_6 with TMS as internal standard. The data are reported as follows: chemical shift in parts per million

(ppm, δ units) and spin-spin coupling *J* (Hz). DMF was dried and distilled over CaH₂, whereas THF was used after distillation over Na/benzophenone.

General Preparation of *N*-(Pg-aminoxyacyl)benzotriazole (2). Thionyl chloride (1.2 mmol) was added to a solution of benzotriazole (4.16 mmol) in anhydrous THF (5 mL) at 0 °C, and the reaction mixture was stirred for 20 min at same temperature. *N*-(Pg)-aminoxyacetic acid (1.0 mol) dissolved in anhydrous THF (3 mL) was added dropwise to the mixture. After being stirred for 4 h at 0 °C, the reaction mixture was allowed to warm to room temperature. After 1 h, the white precipitate was filtered off, and the filtrate was concentrated under reduced pressure. The residue was diluted with CH₂Cl₂ (25 mL), and the solution washed with saturated Na₂CO₃ solution (3 × 10 mL) and then saturated brine solution and finally dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure to afford *N*-(Pg-aminoxyacyl)benzotriazoles (2).

Boc-AOGly-Bt (2a): White microcrystals (67%); mp 115–116 °C; ¹H NMR (CDCl₃) δ 1.50 (s, 9H), 5.54 (s, 2H), 7.54 (t, J = 7.5 Hz, 1H), 7.70 (t, J = 7.8 Hz, 1H), 7.96 (s, 1H), 8.14 (d, J = 8.1 Hz, 1H), 8.27 (d, J = 8.1 Hz, 1H); ¹³C NMR (CDCl₃) δ 28.3, 74.6, 82.7, 114.1, 120.6, 126.8, 130.9, 131.0, 146.0, 156.4, 168.3. Anal. Calcd for C₁₃H₁₆N₄O₄: C, 53.42; H, 5.52; N, 19.17. Found: C, 53.07; H, 5.69; N, 18.59.

General Preparation of N-(Pg)-aminoxy Acid Amides (3). Amine (1 equiv) and triethylamine (1 molar equiv) in THF (2 mL) were added to a stirred solution of N-(Pg-aminoxyacyl)benzotriazole (2) (1 molar equiv) in THF (4 mL) dropwise at 0 °C, and the mixture was stirred for 2 h at room temperature. After evaporation of THF, EtOAc (20 mL) was added to the solution, which was washed with saturated Na₂CO₃ solution (3 × 10 mL) and brine (10 mL) and dried over anhydrous Na₂SO₄. Evaporation of the solvent under reduced pressure gave the crude amides.

tert-Butyl 2-(Isopropylamino)-2-oxoethoxycarbamate (3a)¹³. The crude product was recrystallized from diethyl ether-hexanes to give white microcrystals (74%): mp 67–68 °C; ¹H NMR (CDCl₃) δ 1.19 (d, J = 6.6 Hz, 6H), 1.49 (s, 9H), 4.00–4.20 (m, 1H), 4.27 (s, 2H), 7.51 (s, 1H), 8.00 (s, 1H); ¹³C NMR (CDCl₃) δ 22.5, 28.1, 41.0, 76.1, 83.0, 157.8, 167.8. Anal. Calcd for C₁₀H₂₀N₂O₄: C, 51.71; H, 8.68; N, 12.06. Found: C, 52.09; H, 8.90; N, 12.12.

General Preparation of α -AO- α -hybrid Dipeptides 4 and α -AO- α , α -hybrid Tripeptides 6. The unprotected amino acids (1.5 mmol) and triethylamine (2.0 mmol) were dissolved in a minimum amount of water. Acetonitrile (3 mL) was added, and the solution was cooled to 0 °C. A solution of *N*-(Pg-aminooxya-cyl)benzotriazole (2) (1 mmol) in acetonitrile (4 mL) was added dropwise over 10 min at 0 °C and the resulting solution stirred for 4 h at 10 °C. After evaporation of THF, EtOAc (20 mL) was

added, and the mixture was washed with 4 N HCl solution (3 \times 15 mL) and brine (15 mL) and dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure to give crude product **4** or **6**.

Cbz-AOGly-L-Phe-OH (4a). The residue was purified by column chromatography [EtOAc-hexanes (from 1:3 to 1:1)] to give an oil (51%): $[\alpha]_{23}^{D3} = -9.0 (c \ 1.00, CH_3OH); {}^{1}H \ NMR (CDCl_3) \delta \ 3.02 (dd, J = 14.0 \ Hz, 8.3 \ Hz, 1H), 3.25 (dd, J = 14.0 \ Hz, 4.9 \ Hz, 1H), 4.28 (s, 2H), 4.76-4.89 (m, 1H), 5.11 (s, 2H), 7.14-7.30 (m, 5H), 7.33 (s, 5H), 7.72 (br s, 1H), 8.08 (s, 1H), 8.23 (br s, 1H); {}^{13}C \ NMR (CDCl_3) \delta \ 37.3, 53.6, 68.5, 75.8, 127.2, 128.6, 128.8, 128.9, 129.4, 135.1, 136.2, 158.5, 169.8, 174.5. Anal. Calcd for C₁₉H₂₀N₂O₆.H₂O: C, 58.46; H, 5.16; N, 7.18. Found: C, 58.76; H, 5.42; N, 7.38.$

Cbz-AOGly-Gly-L-Phe-OH (6a). The residue was recrystallized from diethyl ether-hexanes to give white hydroscopic microcrystals, (81%): mp 29–31 °C; $[\alpha]_{23}^{23} = +7.1$ (*c* 1.00, CH₃-OH); ¹H NMR (CDCl₃) δ 2.92 (br s, 1H), 3.04 (br s, 1H), 3.76 (br s, 1H), 3.83 (br s, 1H), 4.22 (br s, 2H), 4.67 (br s, 1H), 5.00 (br s, 2H), 6.92–7.36 (m, 12H), 8.04 (s, 1H), 8.73 (s, 1H); ¹³C NMR (CDCl₃) δ 37.4, 42.8, 53.8, 68.4, 75.9, 127.3, 128.3, 128.6, 128.8, 128.9, 129.6, 135.3, 136.1, 158.6, 169.8, 170.5, 173.8. Anal. Calcd for C₂₁H₂₃N₃O₇: C, 58.74; H, 5.40; N, 9.79. Found: C, 58.57; H, 5.28; N, 9.60.

General Preparation of α,α-AO-hybrid Dipeptides 9. α-Aminoxy acid hydrochloric acid salts 8 (1.5 mmol) and triethylamine (3.5 mmol) were dissolved in minimum amount of water. Acetonitrile (3 mL) was added, and the solution was cooled to 0 °C. A solution of *N*-(Pg-aminoacyl)benzotriazole (7) (1 mmol) in acetonitrile (3 mL) was added dropwise over 10 min at 0 °C, and the resulting solution was stirred for 4 h at 10 °C. The reaction was evaporated under vacuum. The mixture was diluted with EtOAc (20 mL), washed with 4 N HCl solution (3 × 15 mL) and brine (15 mL), and dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure to give crude product 9.

Cbz-L-Ala-AOGly-OH (9a). The residue was purified by column chromatography [EtOAc-hexanes (from 1:3 to 1:1)] to obtain a sticky oil (61%): $[\alpha]_D^{23} = -27.8 (c \ 1.00, CH_3OH)$; ¹H NMR (CDCl₃) δ 1.39 (d, J = 6.9 Hz, 3H), 4.31–4.51 (m, 1H),

4.51 (s, 2H), 5.06 (d, J = 12.3 Hz, 1H), 5.13 (d, J = 12 Hz, 1H), 5.59 (d, J = 8.4 Hz, 1H), 7.26–7.45 (m, 6H), 10.42 (br s 1H); ¹³C NMR (CDCl₃) δ 18.3, 48.0, 68.0, 73.3, 128.3, 128.7, 128.8, 135.6, 156.7, 171.2, 171.9. Anal. Calcd for C₁₃H₁₆N₂O₆: C, 52.70; H, 5.44; N, 9.46. Found: C, 53.08; H, 5.72; N, 9.19.

General Preparation of α -Aminoxy Dipeptides 10. α -Aminoxy acid hydrochloric acid salts (8) (1.5 mmol) and triethylamine (3.5 mmol) were dissolved in a minimum amount of water. Acetonitrile (3 mL) was added, and the solution was cooled to 0 °C. A solution of *N*-(Pg-aminoxyoacyl)benzotriazole (2) (1 mmol) in acetonitrile (3 mL) was added dropwise over 10 min at 0 °C, and the resulting solution was stirred for 4 h at 10 °C. After the solvent was evaporated under vacuum, EtOAc (20 mL) was added, and the mixture was washed with 3 N HCl solution (3 × 15 mL) and brine (15 mL) and dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure to give crude product 10.

Cbz-L-AOVal-AOGly-OH (10a). The residue was an oily mixture of 10a (80%) and 1f (20%): $[\alpha]_D^{23} = -56.0$ (*c* 1.00, CH₃OH); (data from the mixture) ¹H NMR (CDCl₃) δ 0.91 (d, J = 6.9 Hz, 3H), 1.04 (d, J = 6.9 Hz, 3H), 2.18–2.30 (m, 1H), 4.19 (d, J = 4.2 Hz, 1H), 4.43 (d, J = 17.4 Hz, 1H), 4.54 (d, J = 17.4 Hz, 1H), 5.17 (d, J = 11.7 Hz, 1H), 5.24 (d, J = 12.0 Hz, 1H), 7.32–7.42 (m, 5H), 8.17 (s, 1H), 11.64 (br s, 1H); ¹³C NMR (CDCl₃) δ 16.7, 18.9, 30.9, 69.0, 75.1, 91.2, 128.6, 128.7, 128.8, 128.9, 129.0, 134.7, 159.5, 171.1, 172.0; HRMS [M + Na]⁺ found 363.1175, theoretical for C₁₅H₂₀N₂O₇·Na⁺ 363.1163.

Acknowledgment. We thank Dr. C. D. Hall for helpful discussions.

Supporting Information Available: Materials and methods; general procedure for preparation of *N*-protected aminoxy acids 1b-g and their characterization data; characterization data of 2b-2g, 3b-g, (3g+3g'), (4a+4a')-4h, 6b-d, (9a+9a')-9e, and 10b, ¹H NMR, ¹³C NMR spectra of compounds and chiral HPLC chromatograms. This material is available free of charge via the Internet at http://pubs.acs.org.